## **Supporting Information**

# Supercoiling Structure-based Design of a Trimeric Coiled-coil Peptide with High Potency against HIV-1 and Human $\beta$ -coronavirus Infection

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Table S1. Summary of the SVA results of N3G alone, N3G/C34 and N3G/HR2P complexes<sup>a</sup>

Peptides	Sedimentation coefficient (s)	Observed molecular mass (kDa)	Calculated molecular mass (kDa)
N3G	0.77	12.8	13.2
N3G/C34 6-HB	2.15	25.3	26.1
N3G/MERS-CoV HR2P 6-HB	2.13	23.2	25.8
N3G/HCoV-OC43 HR2P 6-HB	2.08	23.4	26.2

<sup>&</sup>lt;sup>a</sup> SVA studies were performed at a concentration of 150 μM in PBS (pH 7.4) and a rotor speed of 60,000 rpm at 20 °C.

**Table S2.** Biophysical properties of the designed peptides

Compd	Helicity (%) <sup>a</sup>	$Tm ( \mathfrak{C})^b$
N3G	69.5	>90
N1G	74.7	55.0
N3HR	74.8	>90

<sup>&</sup>lt;sup>a</sup> CD spectra of each designed peptide were monitored in PBS, pH 7.4. The final concentration of the peptide was 15 μM.

**Table S3.** HPLC method used for the purification of peptide compounds<sup>a</sup>

Time (min)	Solvent A (%)	Solvent B (%)
5	70	30
13	53	47
30	17	83
35	17	83
55	70	30
60	70	30

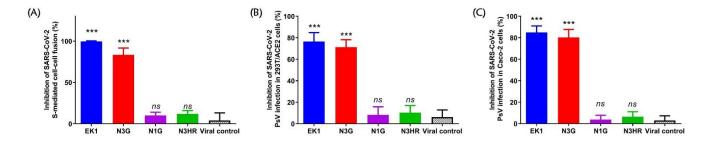
<sup>&</sup>lt;sup>a</sup> The crude peptide products were purified by preparative reverse phase HPLC with a Waters preparative HPLC system (PrepLC 4000) on a Waters X-bridge C8 column (19.5mm  $\times$  250mm, 10μm) at constant flow rate of 16 mL/min. Solvent A: 0.1% trifluoroacetic acid in H<sub>2</sub>O; Solvent B: 0.1% trifluoroacetic acid in 70% CH<sub>3</sub>CN/H<sub>2</sub>O.

Table S4. HPLC method used for the analysis of peptide compounds<sup>a</sup>

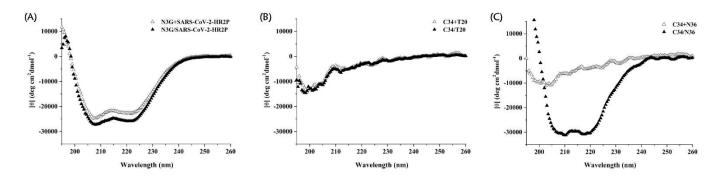
Methods	Time (min)	Solvent A (%)	Solvent B (%)
	5	40	60
Method A	12	10	90
Method A	20	0	100
	25	90	10
Method B	5	50	50
	10	0	100
	20	0	100
	25	90	10

<sup>&</sup>lt;sup>a</sup> Peptide compounds were analyzed by analytical RP-HPLC performed on a RP-C8 column (Zorbax Eclipse XDB-C8,  $4.6 \times 150$  mm, 5 μm) using two different solvent systems (Methods A and B) and a flow rate of 1 mL/min with detection at 210 nm. Solvent A: 0.1% trifluoroacetic acid in H<sub>2</sub>O; Solvent B: 0.1% trifluoroacetic acid in 70% CH<sub>3</sub>CN/H<sub>2</sub>O.

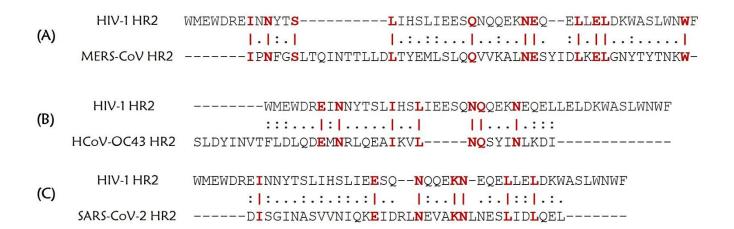
<sup>&</sup>lt;sup>b</sup> Thermal denaturation CD scans were performed on 15 μM peptide solutions in PBS with 2 M guanidine hydrochloride.



**Figure S1.** Both N3G and EK1 are effective against SARS-CoV-2 infection. (**A**) Inhibition of peptides on SARS-CoV-2 S-mediated cell–cell fusion at the screening concentration of 20  $\mu$ M. Inhibition of peptides against pseudotyped SARS-CoV-2 infection in 293T/ACE2 cells (**B**) and Caco-2 cells (**C**). Data are presented as mean  $\pm$  s.d. of three independent experiments. \*\*\* indicates a significant difference from viral control (Student's *t* test, P < 0.001). ns: not significant.



**Figure S2.** N3G and SARS-CoV-2-HR2P show interaction in solution. (**A**) CD spectrum of SARS-CoV-2-HR2P/N3G mixtures (solid symbol) and the theoretical noninteracting spectra of the related isolated peptides (SARS-CoV-2-HR2P+N3G, open symbol) are shown for comparison. N3G and SARS-CoV-2-HR2P interaction induces more α-helix structure than the sum of the single peptides, which is distinguishable from (**B**) noninteracting T20-C34 peptide pair. (**C**) As a positive control, the interaction between N36 and C34 led to their dramatic secondary structural changes. The final concentration of each peptide in PBS (pH 7.4) was 15 μM.



**Figure S3.** Sequence similarity between the HR2 domain (residues 628–673) in gp41 of HIV-1 and the N-terminal portion of that of (**A**) MERS-CoV (residues 1,246–1,295), (**B**) HCoV-OC43 (residues 1,246–1,295), or (**C**) SARS-CoV-2 (residues 1,168–1,203). Identical amino acid residues are highlighted in red.

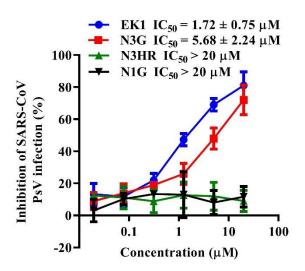


Figure S4. Inhibitory activity of peptides on pseudotyped SARS-CoV infection in Caco-2 cells.

## MALDI-TOF-MS and analytical HPLC of designed peptides

